

# Monkeypox Specimen Submission Instructions for Providers



## Monkeypox Testing Authorization

Clinicians requesting testing for monkeypox must first complete the Indiana Department of Health (IDOH) [Monkeypox Specimen Authorization Request form](#).

Once approved, specimens should be submitted via [LimsNet](#), an online system that will make results available as PDF files the minute they are released at the lab. Specimens should be submitted through the Biothreat submission page in LimsNet. **Each set of lesion swabs (2 swabs per one lesion) should be submitted separately in LimsNet, creating up to two submissions per one patient.** To get a free LimsNet account established at your facility, call the help desk at (317) 921-5506 or email [LimsAppSupport@isdh.in.gov](mailto:LimsAppSupport@isdh.in.gov). Guidance on how to submit specimens in LimsNet is available [here](#).

## Monkeypox Testing Process

Testing for monkeypox includes preliminary testing for non-variola orthopoxvirus at the Indiana Department of Health Laboratories (IDOHL), followed by confirmatory monkeypox testing at the Centers for Disease Control and Prevention (CDC). In the current outbreak, orthopoxvirus positive results are presumed to be monkeypox unless shown to be otherwise by CDC testing. Only specimens positive for orthopoxvirus at the IDOHL will be sent to CDC for confirmatory testing.

## Turnaround Time and Results

Typical turnaround time for results at the IDOHL is typically within a day of specimen receipt at IDOHL. Results will be released directly to submitters via LimsNet. **Providers are responsible for relaying results to the patient.**

## Specimen Collection Instructions

Personnel who collect specimens for monkeypox testing should use personal protective equipment (PPE) in accordance with CDC's [recommendations for healthcare settings](#).

Providers should collect swabs from more than one lesion, preferably from different locations on the body and/or from lesions with differing appearances. Providers should swab the lesion vigorously to collect adequate DNA; it is not necessary to de-roof the lesion before swabbing. Separate swabs must be used for each lesion, and two swabs must be submitted for each lesion sampled (one for preliminary and one for confirmatory testing). No more than two lesion sites (four swabs total) may be submitted for each patient. Specimens must be clearly labeled with the site the specimen was collected from, patient name, date of birth, and date of specimen collection.

## Specimen Collection Materials and Storage

Dry synthetic swabs must be used for collection. Larger swabs (such as those used for oropharyngeal specimens) are preferred, but swabs for COVID-19 or flu testing may be used if no other swabs are available. Swabs should be placed into 1.5- or 2-mL sterile, screw-capped tubes with O-ring or 15-mL sterile, screw-capped tubes. If your facility does not have these tubes available, any sterile, screw-capped tube (such as a TB sputum cup or sterile urine cup) may be used. **The specimen must be a dry swab. Do not use any transport media. Specimens must be stored at refrigerated temperatures within 1 hour after specimen collection. Specimens may be frozen if longer storage is required.**

## Specimen Submission

Once collected, specimens should be shipped using **Category B** shipping materials. Specimens **MUST** be placed in a Styrofoam container with **sufficient cold packs to maintain refrigerated temperatures during transport**. Specimens should be shipped on dry ice if frozen or if transit time will be longer than 24 hours. Specimens must be shipped to arrive **COLD** at the IDOH Laboratories between 8:15 AM and 4:45 PM Monday – Friday. **Do not ship a specimen that will be transported over multiple days or the weekend (e.g., shipping on Friday for Monday delivery).**

## Questions

For questions related to monkeypox testing, please contact the IDOH Epidemiology Resource Center at 317-233-7125 during normal business hours (8:15 AM – 4:45 PM, M-F) or 317-233-1325 after hours, weekends, or holidays.

<b>Title</b>	Non-variola Orthopoxvirus Specimen Collection and Transport
<b>Specimen Requirements</b>	<ol style="list-style-type: none"> <li>1. Dry swab of lesion is preferred.</li> <li>2. Vesicle fluid, skin, crust, "roof"</li> </ol>
<b>Sampling Materials</b>	<ol style="list-style-type: none"> <li>1. Swab – Swab material should be synthetic, i.e., rayon, polyester, or Dacron. Calcium alginate or charcoal-impregnated swabs should not be used, nor should wood-shaft swabs.</li> <li>2. 1.5- or 2-mL sterile, screw-capped tube with O-ring or 15-mL sterile, screw-capped tube.</li> <li>3. For collection of vesicle fluid, a 26 or 27-gauge needle attached to a tuberculin syringe is necessary.</li> <li>4. Cold packs or dry ice.</li> <li>5. Shipping boxes/containers with appropriate shipping labels (Category B Infectious Substance).</li> </ol>
<b>Procedural Notes</b>	<ol style="list-style-type: none"> <li>1. Be sure to properly label the specimen tube with at least the patient's name and date of collection.</li> <li>2. After collection, all specimens should be stored at refrigerated temperature (2-8°C) until shipped. If longer storage is required, specimens can be frozen.</li> <li>3. Special Instructions for Specimen Collection <ul style="list-style-type: none"> <li>• Vigorously swab or brush lesion with two separate sterile dry polyester or Dacron swabs;</li> <li>• Break off end of applicator of each swab into a 1.5- or 2-mL screw-capped tube with O-ring or place each entire swab in a separate sterile container (15-mL sterile, screw-capped tube). Do not add or store in viral or universal transport media.</li> </ul> </li> <li>4. Specimens should be submitted by LimsNet, an online system that will make results available as PDF files the minute they are released at the lab. Submit through the Biothreat submission page in LimsNet.</li> <li>5. To get a free LimsNet account established at your facility for electronic submission and results reporting, call the help desk at (317) 921-5506 or email to <a href="mailto:LimsAppSupport@isdh.in.gov">LimsAppSupport@isdh.in.gov</a>.</li> <li>6. Alternatively, a completed Clinical Biothreat Agent Submission Form must accompany each specimen. Contact the IDOHL BT-Clinical Microbiology division director at (317) 921-5842.</li> </ol>
<b>Shipping Instructions</b>	<ol style="list-style-type: none"> <li>1. Wrap the labeled specimen container with absorbent material and place in a biohazard specimen bag. Be sure to package each patient's specimens individually to avoid cross-contamination.</li> <li>2. Place the requisition form in the side pocket of the biohazard bag. Never place the requisition form in with the specimen in case the specimen leaks during transit. If the specimen bag does not have 2 compartments, place the paperwork in a separate ziploc bag.</li> <li>3. Place the specimen(s) in a styrofoam container with sufficient cold packs to maintain refrigerated temperature during shipment. For swabs, if needed, dry ice can also be used if the specimen is frozen and/or transport time may be longer than 24 hours. If dry ice is used, do not form an airtight seal on the styrofoam container because dry ice releases carbon dioxide gas.</li> <li>4. Place the styrofoam container into a cardboard shipping box, close lid, and seal.</li> <li>5. Ship or transport by courier, compliant with DOT and IATA regulations (Category B Infectious Substance). <ol style="list-style-type: none"> <li>a. Ship to: Indiana Department of Health Laboratories Biothreat Laboratory 550 W. 16th Street, Suite B Indianapolis, IN 46202</li> </ol> </li> </ol>
<b>Reporting and TAT</b>	<p>1-2 business days.</p> <p>Test Referral. Specimens may be forwarded to the CDC for additional testing.</p>